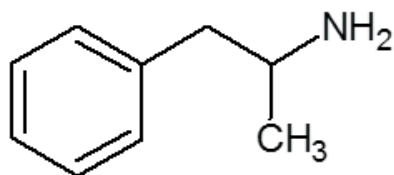


# Extraction of Amphetamines and Analogues from Hair Using ISOLUTE® SLE+ Prior to GC/MS Analysis



**Figure 1.** Structure of amphetamine.

## Introduction

This application note describes the extraction of amphetamine and amphetamine-style compounds from hair matrix prior to GC/MS analysis following bead homogenization.

This application note describes an effective and efficient ISOLUTE® SLE+ protocol optimized for 1 mL capacity formats. The simple sample preparation procedure delivers clean extracts and analyte recoveries greater than 72% with RSDs lower than 10% for all analytes.

In addition, this note describes how to achieve quantitation of all analytes down to 0.2 ng/mg of hair. This complies with the LOQ values for laboratories involved in amphetamine testing, as suggested by the Society of Hair Testing (SoHT).

ISOLUTE SLE+ Supported Liquid Extraction plates and columns offer an efficient alternative to traditional liquid-liquid extraction (LLE) for bioanalytical sample preparation, providing high analyte recoveries, no emulsion formation, and significantly reduced sample preparation.

## Analytes

Amphetamine, Methamphetamine, MDA, MDMA, MDEA, Amphetamine-D<sub>5</sub> (internal standard).

## Sample Preparation Procedure

### Format

ISOLUTE® SLE+ 1 mL Sample Volume Columns, part number 820-0140-C.

### Sample Pre-treatment

Transfer 20 mg of hair into a 2 mL Lysera tube containing five 2.8 mm ceramic beads. Add methanol (1 mL) and internal standard solution (20 µL) before sealing the tube tightly. Homogenize the sample using the Biotage® Lysera 24 for four cycles of 3 minutes at 5.3 m/s with a dwell time of 0.2 minutes. Centrifuge the sample for 10 minutes at 13,300 rpm.

Add 20 µL of ethylene glycol to a clean glass tube and transfer 0.7 mL of supernatant sample to this new tube prior to evaporation at 20 °C. The glycol prevents full evaporation and assists against the loss of amphetamine and methamphetamine.

Reconstitute the dried sample in 0.1% ammonium hydroxide (aq) (850 µL) – see notes for preparation.

### Sample Loading

Load the pre-treated hair extract (0.8 mL) onto the column and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

### Analyte Extraction

Apply MTBE (3 mL) and allow to flow under gravity for 5 minutes. Apply a further aliquot of MTBE (3 mL) and allow to flow for another 5 minutes under gravity. Apply vacuum or positive pressure to pull through any remaining extraction solvent. (5–10 seconds).

Collect in an appropriate glass tube with 100 µL HCl in methanol (0.05 M – see notes for preparation). This acts to stabilize free-base analytes in the solvent prior to evaporation.

### Post Elution and Reconstitution

Dry the extract in a stream of air or nitrogen using a Biotage® SPE Dry at ambient temperature, 20 to 40 L/min, or a TurboVap® at ambient temperature, 1.5 L/min, for 20 minutes.

Reconstitute dried samples with 50 µL ethyl acetate and 50 µL pentafluoropropionic anhydride (PFPA). Vortex-mix and place on a heating block for 15 minutes at 50 °C for optimum amphetamine results.

Evaporate the extract in a stream of air or nitrogen using a SPE Dry (ambient room temperature, 20 to 40 L/min).

Reconstitute extracts with 25 µL ethyl acetate and vortex.

## GC Conditions

### Instrument

Agilent 7890A with QuickSwap

### Column

Restek RXi-5ms, 30 m x 0.25 mm ID x 0.25 µm)

### Carrier

Helium 1.2 mL/min (constant flow)

### Inlet

250 °C, Splitless, purge flow: 50 mL/min at 1.0 min

### Injection

2 µL

### Wash Solvents

Methanol and ethyl acetate

### Oven

Initial temperature 60 °C

Ramp 25 °C/min to 260 °C

### Post Run

Backflush for 1.6 minutes (2 void volumes)

### Transfer Line

280 °C

## Mass Spectrometry Conditions

### Instrument

Agilent 5975C

### Source

230 °C

### Quadrupole

150 °C

### MSD Mode

SIM

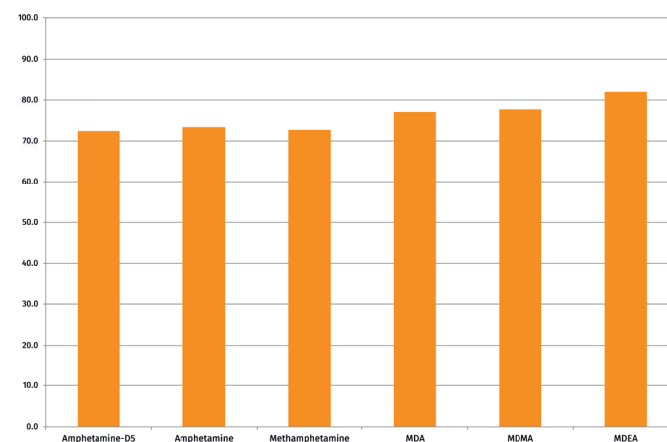
### SIM Parameters

**Table 1.** Ions acquired in the Selected Ion Monitoring (SIM) mode.

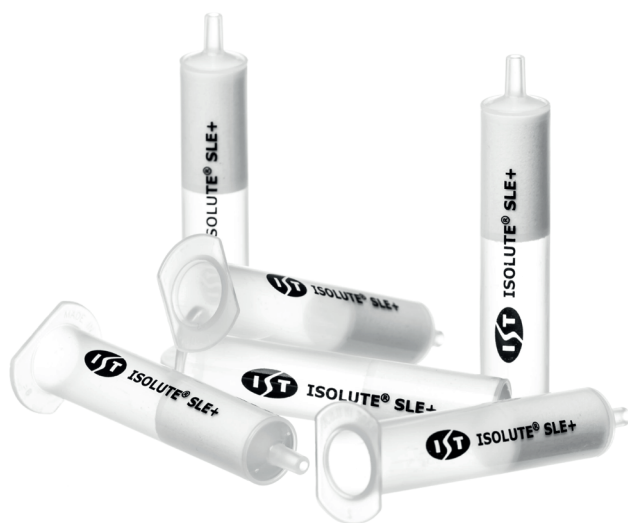
SIM Group	Analyte	Target (Quant) Ion	1st Qual Ion	2nd Qual Ion
1	Amphetamine-D <sub>5</sub>	194	96	123
1	Amphetamine	190	91	118
2	Methamphetamine	204	118	160
3	MDA	162	135	
4	MDMA	162	204	
4	MDEA	218	162	

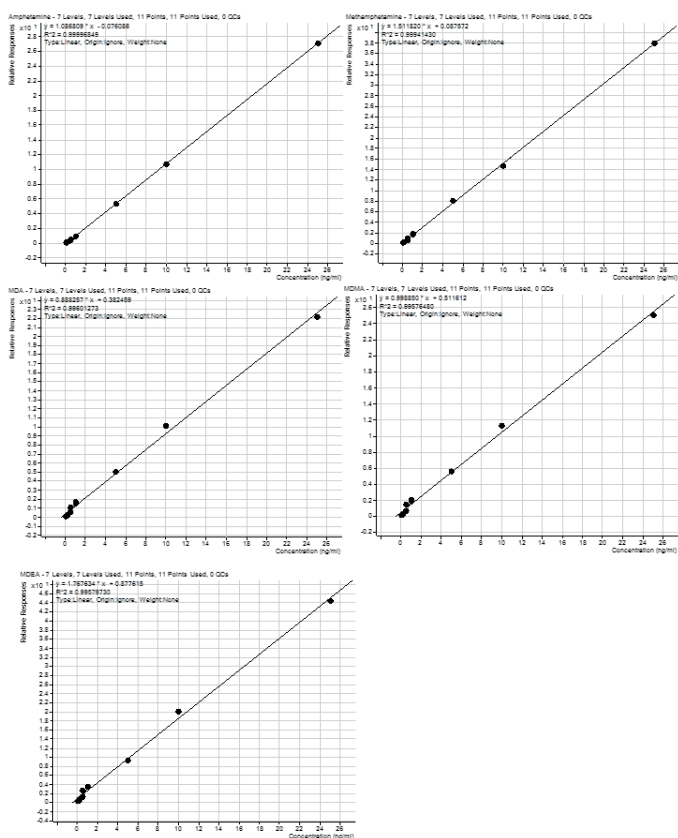
## Results

This optimized ISOLUTE® SLE+ protocol demonstrated analyte recoveries greater than 70% as shown in **Figure 2**. RSDs were below 10% for all analytes.



**Figure 2.** Typical analyte % extraction recoveries (n=7) using the ISOLUTE® SLE+ Protocol.





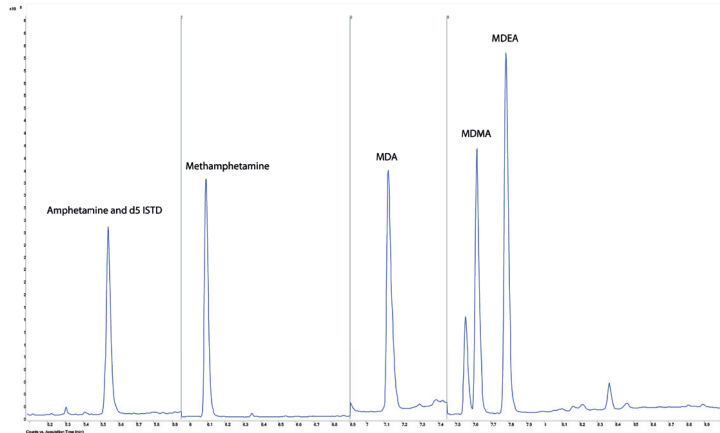
**Figure 3.** Calibration curves for extracted levels of spiked homogenized hair matrix, using 1 mL ISOLUTE® SLE+ format. Concentrations are 0.1, 0.2, 0.5, 1, 5, 10 and 25 ng/mg showing  $R^2$  values of 0.9957 to 0.9999.

## Additional Information

### Reagent Preparation

**0.1% ammonium hydroxide** is prepared by adding 50  $\mu$ L concentrated ammonium hydroxide to 49.95 mL of HPLC grade water. Concentrated stock used to modify pH prior to extraction is commercially available 28–30%.

**0.05 M HCl in methanol** is prepared by adding 50  $\mu$ L concentrated hydrochloric acid to 11.95 mL of HPLC grade methanol. Concentrated stock used to modify pH prior to extraction is commercially available 12M.



**Figure 4.** Representative chromatography for application analytes spiked at 75 ng/mg.

**Table 2.** Lower Limits of Quantitation (LLOQ) using the optimized ISOLUTE® SLE+ procedure.

Drug Analyte	LLOQ (ng/mg)
Amphetamine	0.2
Methamphetamine	0.2
MDA	0.2
MDMA	0.2
MDEA	0.2

## Ordering Information

Part Number	Description	Quantity
<b>820-0140-C</b>	ISOLUTE® SLE+ 1 mL Sample Volume Column	30
<b>19-060</b>	Biotage® Lysera*	1
<b>PPM-48</b>	Biotage® PRESSURE+ 48 Positive Pressure Manifold	1
<b>SD-9600-DHS-EU</b>	Biotage® SPE Dry 96 Sample Evaporator 220/240V	1
<b>SD-9600-DHS-NA</b>	Biotage® SPE Dry 96 Sample Evaporator 100/120V	1
<b>415000</b>	TurboVap® LV	1

\*Biotage® Lysera is available in North America, Europe and China only

### EUROPE

Main Office: +46 18 565900  
Toll Free: +800 18 565710  
Fax: +46 18 591922  
Order Tel: +46 18 565710  
Order Fax: +46 18 565705  
order@biotage.com  
Support Tel: +46 18 56 59 11  
Support Fax: +46 18 56 57 11  
eu-1-pointsupport@biotage.com

### NORTH & LATIN AMERICA

Main Office: +1 704 654 4900  
Toll Free: +1 800 446 4752  
Fax: +1 704 654 4917  
Order Tel: +1 704 654 4900  
Order Fax: +1 434 296 8217  
ordermailbox@biotage.com  
Support Tel: +1 800 446 4752  
Outside US: +1 704 654 4900  
us-1-pointsupport@biotage.com

### JAPAN

Tel: +81 3 5627 3123  
Fax: +81 3 5627 3121  
jp\_order@biotage.com  
jp-1-pointsupport@biotage.com

### CHINA

Tel: +86 21 2898 6655  
Fax: +86 21 2898 6153  
cn\_order@biotage.com  
cn-1-pointsupport@biotage.com

### KOREA

Tel: +82 31 706 8500  
Fax: +82 31 706 8510  
korea\_info@biotage.com  
kr-1-pointsupport@biotage.com

Part Number: AN885.V.1

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